



¹ Center of Excellence for Regenerative Dentistry, Faculty of Dentistry, Chulalongkorn University, Bangkok 10330 THAILAND

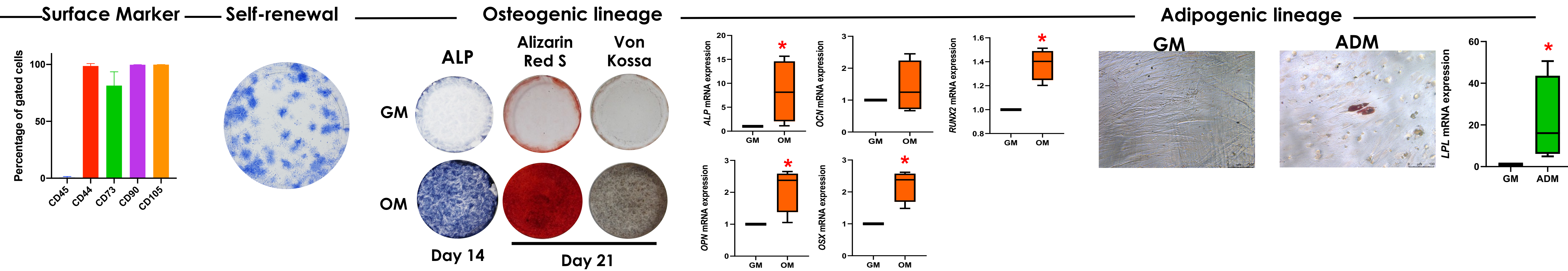
² Centre de Recherche des Cordeliers, Université de Paris, Sorbonne Université, INSERM UMRS 1138, Molecular Oral Pathophysiology; Paris, F-75006, France

³ Institut Curie, PSL Research University, Laboratoire de Spectrométrie de Masse Protéomique, 26 rue d'Ulm, Paris 75248, France

⁴ Genomics and Precision Dentistry Research Unit and Department of Anatomy, Faculty of Dentistry, Chulalongkorn University, Bangkok 10330 THAILAND

⁵ Université de Paris, Dental Faculty Garancière, Paris, F-75006, France

Dental pulp cells exhibited the characteristic of mesenchymal stem cells



Extracellular matrix production and decellularization

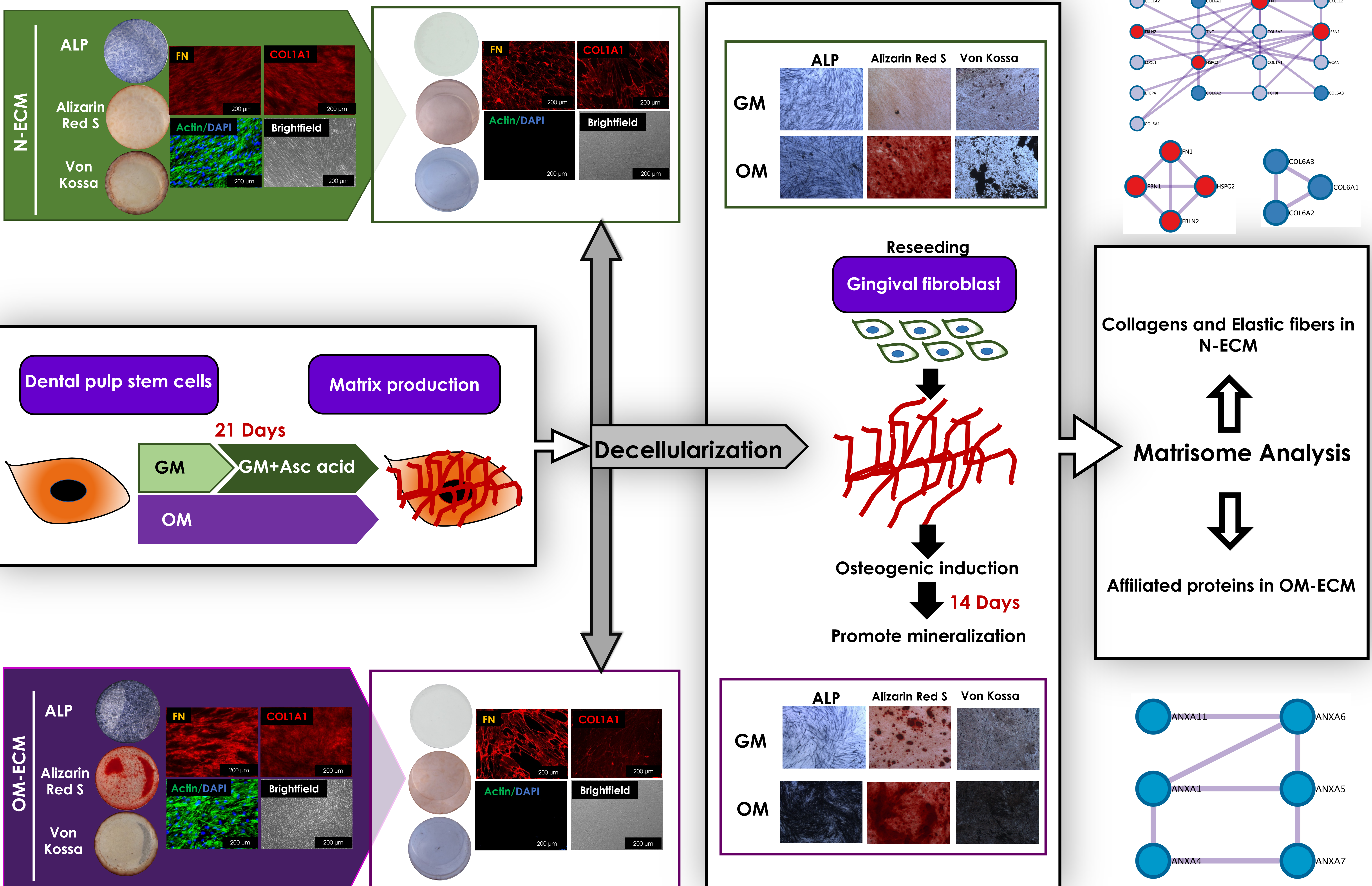
ECM were extracted from DPSCs cultured in normal growth medium supplemented with L-ascorbic acid (N-ECM) or in osteogenic induction medium (OM-ECM). ECM decellularization (dECM) was performed using 0.5% triton X-100 in 20mM ammonium hydroxide after 21 days. Mass spectrometry and proteomic analysis identified and quantified matrisome proteins.

Decellularized extracellular matrix enhanced osteogenic differentiation and mineralization

GF were reseeded on N-dECM and OM-dECM and cultured in normal or osteogenic medium. GF were able to attach and proliferate on N-dECM and OM-dECM. Both dECM enhanced mineralization of GF at day 14 compared to tissue culture plate (TCP). In addition, OM-dECM promoted higher mineralization of GF than N-dECM although cultured in growth medium.

Matrisome analysis of extracellular matrix

Extracellular matrix derived from dental pulp stem cells exhibited specific matrisome protein of elastic fiber in N-ECM and affiliated proteins in OM-ECM.



Conclusion

Matrisome proteins of DPSCs derived ECM differ according to culture conditions, with increased core matrisome proteins (collagens, elastic fibres associated) in N-ECM while matrisome associated proteins (annexins) were enriched in OM-ECM. We demonstrated that ECM impacts cell behavior and differentiation. DPSCs ECM proved to be mineralization inductive. Our identified ECM proteins can be used as a marker for osteogenic differentiation. Further study will investigate the patterns of those proteins during osteogenic differentiation. This *in vitro* ECM or some of its proteins may be helpful for tissue engineering and could be used for mineralized tissue therapy or to decorate biomaterials.

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